Phylogenetic Comparative Methods: I

Phenotypic Diversity

Macroevolutionary patterns of phenotypic diversity through time

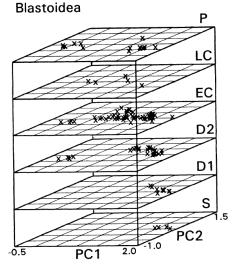
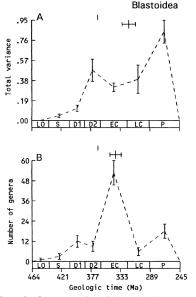
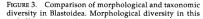
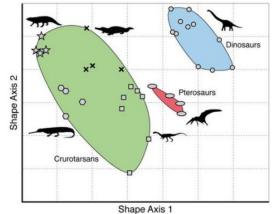


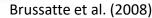
FIGURE 2. Temporal pattern of morphospace occupation in Blastoidea. Note overall increase in range of morphospace occupied. Early Carboniferous taxonomic di-

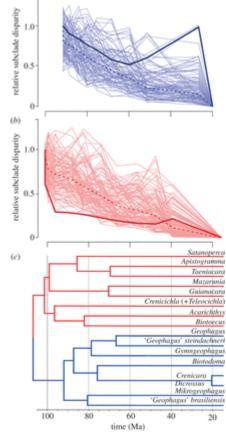
Foote (1993)







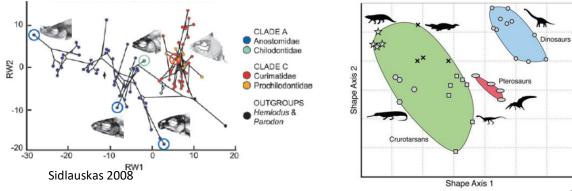




Arbour and López-Fernández (2013)

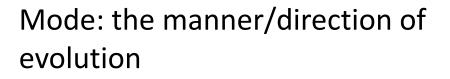
Accumulation of Phenotypic Diversity

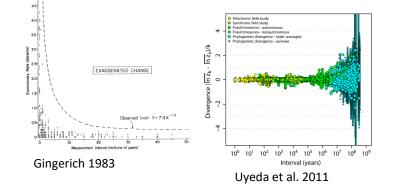
Morphological diversity

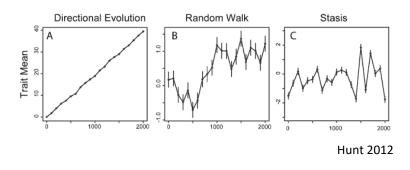


Brussatte et al. 2008

Tempo: the pace of evolution



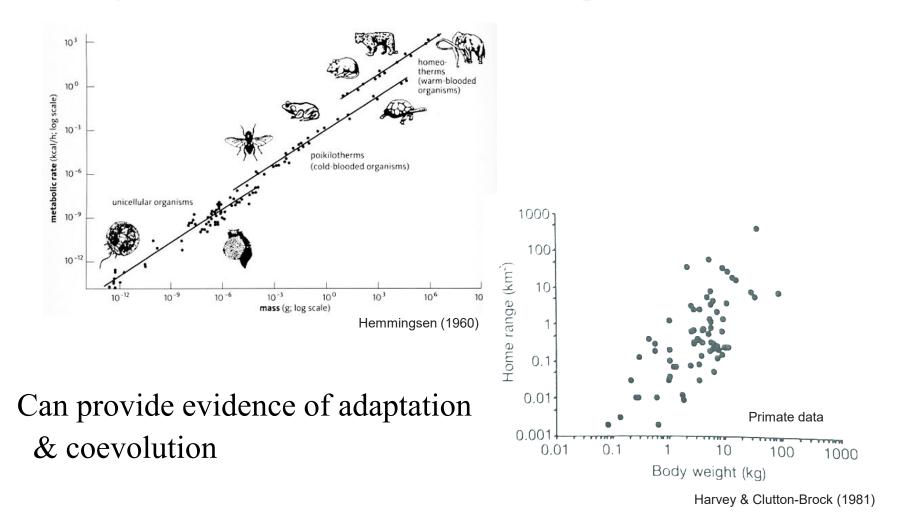




• How do we characterize patterns, and hypothesize processes?

Comparative Evolutionary Biology

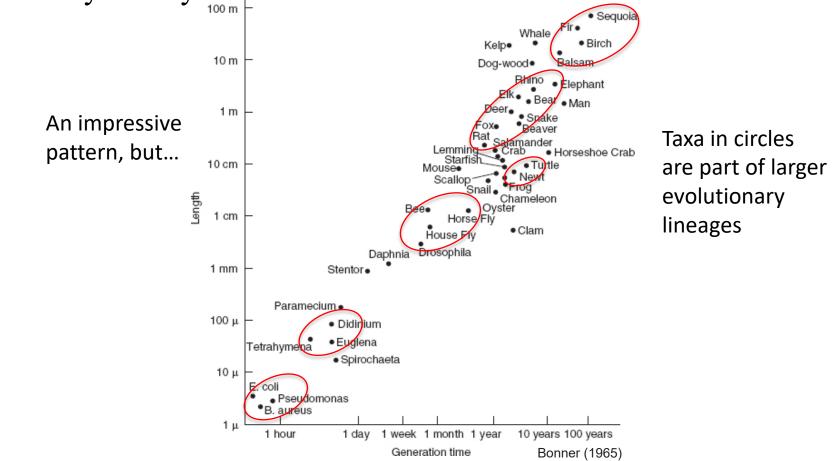
One way: examine trait correlations across species



Unfortunately such comparisons often lead us astray

Comparative Evolutionary Biology

Species-level analyses (or analyses of higher-taxonomic groups) ignore evolutionary history



How many 'independent' data points do we have here??? This can make an enormous difference in inference and interpretation!

Cross-Species Correlations

Say we observe the following pattern

Species	Behavior	Coloration			
1	Solitary 🔵	Cryptic			
2	Solitary 🔵	Cryptic			
3	Solitary 🔵	Cryptic			
4	Solitary 🔵	Cryptic			
5	Solitary 🔵	Cryptic			
6	Solitary 🔵	Cryptic			
7	Solitary 🔵	Cryptic			
8	Solitary 🔵	Cryptic			
9	Gregarious	Aposematic			
10	Gregarious	Aposematic 📃			
11	Gregarious	Aposematic			
12	Gregarious	Aposematic 📃			
13	Gregarious	Aposematic		Solitary	Gregarious
14	Gregarious	Aposematic 📃			_
15	Gregarious	Aposematic	Cryptic	8	0
16	Gregarious	Aposematic 📃	Aposematic	0	8

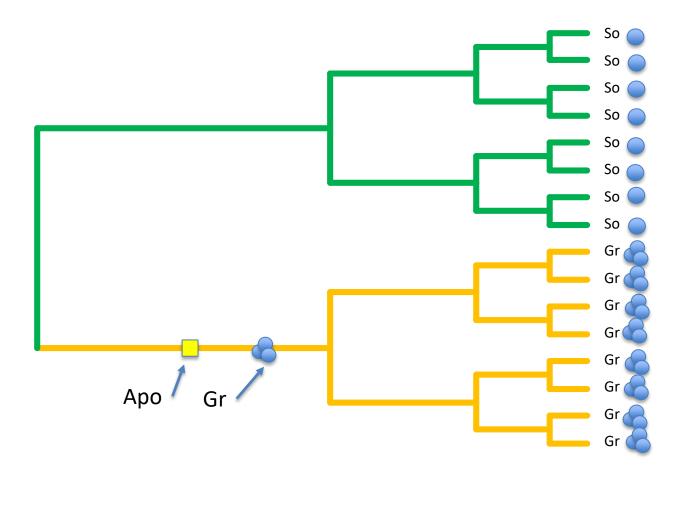
X² = 12.25; *P* = 0.0004

Pretty impressive association!

Inspired by data of Sillen-Tullberg (1988)

Differing Evolutionary Interpretations

But what if evolutionarily we have:

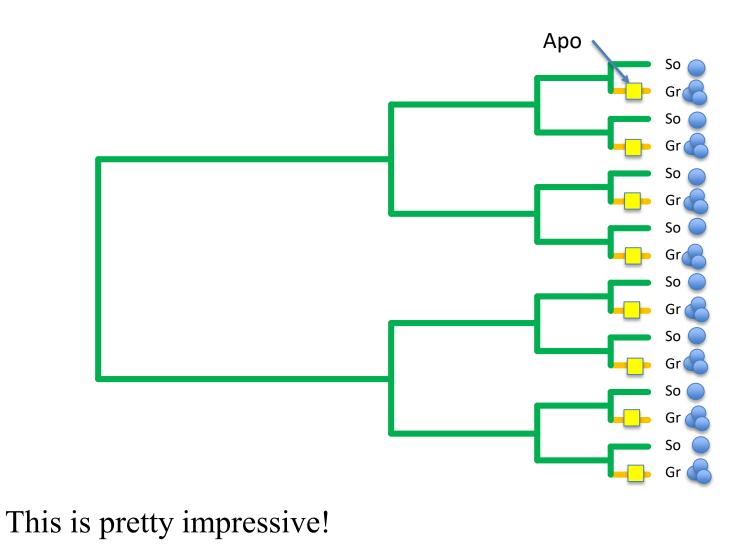


ONE evolutionary change can explain pattern

Not so impressive any longer!

Differing Evolutionary Interpretations

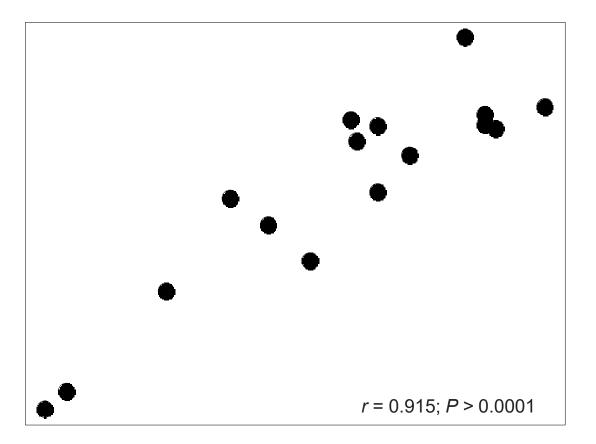
Or what if evolutionarily we have:



Eight correlated evolutionary changes of both traits!

Cross-Species Correlations

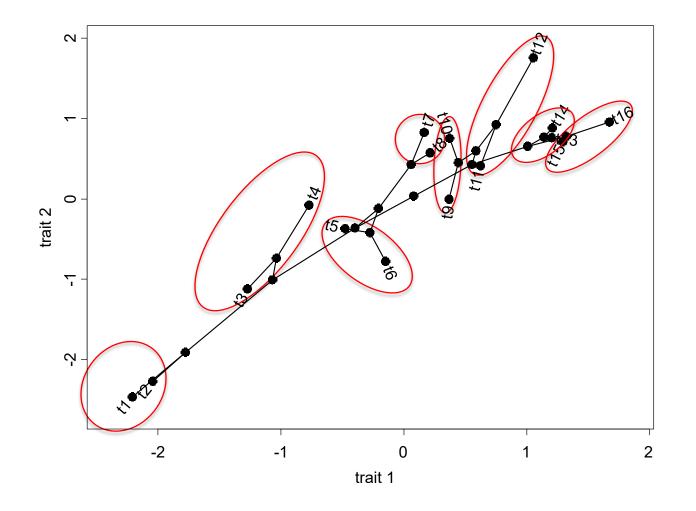
Or what if we observe this pattern:



An impressive association!

Differing Evolutionary Interpretations

But what if evolutionarily we have:

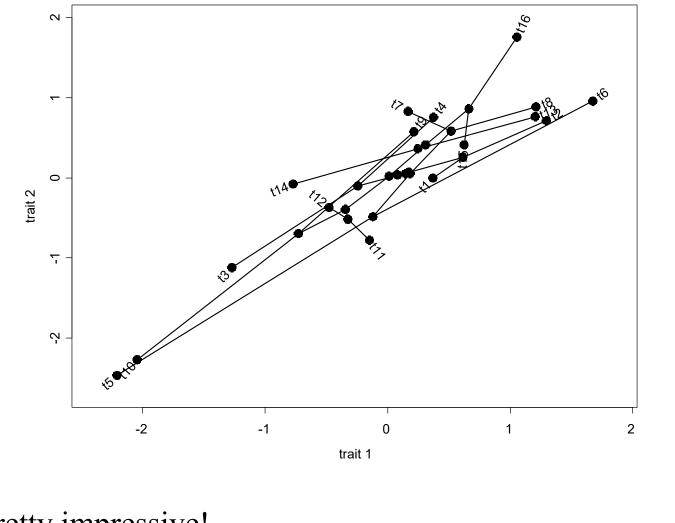


Traits seem to change with phylogeny

Not so impressive, closely related species are similar!

Differing Evolutionary Interpretations

Or what if evolutionarily we have:



Lots of correlated evolutionary change in both traits

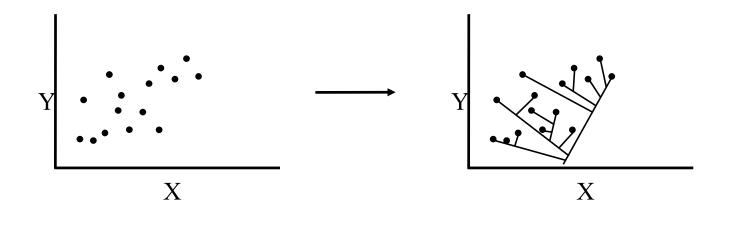
Still pretty impressive!

Comparative Evolutionary Biology

The point:

-Taxa are not independent

-Ignore evolutionary history AT ONE'S PERIL!



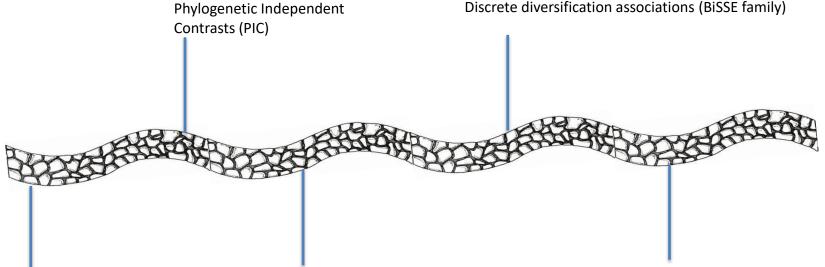
Phylogenetic comparative methods <u>*condition*</u> the data on the phylogeny to account for lack of independence during the analysis

Phylogenetic Comparative Methods

An Incomplete History



Synthesis: PIC, PGLS, Phylo-transform Complex model comparison (BM1, BMM, OU1, OMM) Bayesian methods Parameter-shift methods (e.g., MEDUSA, BAMM) Discrete diversification associations (BiSSE family)



70s – early 80s: early attempts

Nested ANOVA Phylogenetic autocorrelation Discrete change correlations

80s - 90s: 'niche expansion'

1985: The Breakthrough

PGLS Phylogenetic signal (λ , K) Phylogenetic ANOVA Evolutionary models (BM1, OU1, ACDC, λ) Diversity plots (LTT & DTT) Diversification rates Discrete trait change models 2010s: Multivariate approaches

TODAY: PCMs: A diverse toolkit for evaluating evolutionary hypotheses

Comparative Methods: Discrete Data

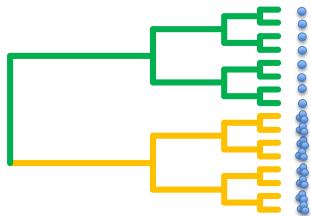
Testing association of discrete traits without phylogeny is problematic

Species	Behavior	Coloration	
1	Solitary 🔵	Cryptic	
2	Solitary 🔵	Cryptic	
15	Gregarious	Aposematic 📃	
16	Gregarious	Aposematic 📃	

Tally of co-occurrence of states ignores number of *INDEPENDENT EVOLUTIONARY TRANSITIONS*

	Solitary	Gregarious
Cryptic	8	0
Aposematic	0	8

X² = 12.25; *P* = 0.0004



But what if association of traits not independent of phylogeny?

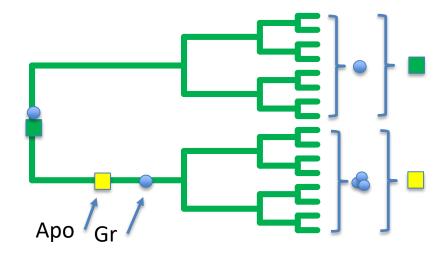
Discrete Data: Counting Evolutionary Events

Identify the number of evolutionary *changes* in each trait

-Associate changes with one another

Procedure:

- -Map traits to phylogeny
- -Estimate ancestral states*
- -Tally transitions in each trait
- -X² test for significance



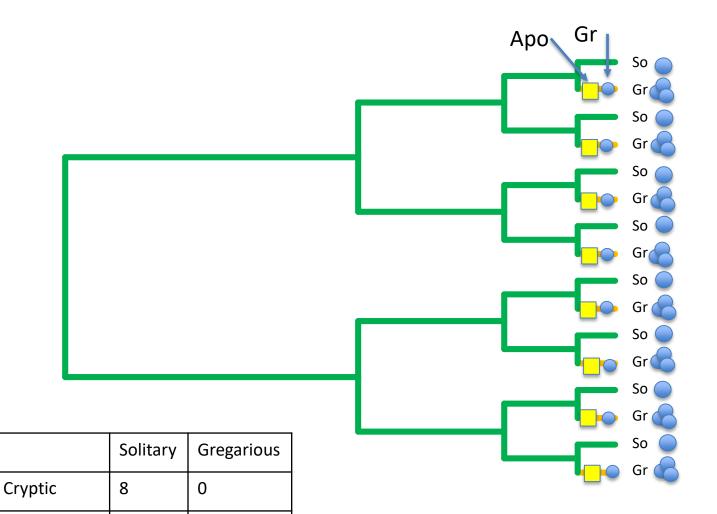
NOTE: method counts transitions, but not direction of change

	Solitary	Gregarious
Cryptic	1	0
Aposematic	0	1

 $X^2 = 0.00; P = 1.00$

Ridley (1983) proposed outgroup comparison: more sophisticated methods exist

Discrete Data: Counting Evolutionary Events Another example:



X² = 12.25; *P* = 0.0004

Aposematic

0

8

Discrete Data: Concentrated Changes

Problem: counting evolutionary events (ala Ridley 1983) does not consider the distribution of changes in one trait *relative* to changes in the other

CCT: Evaluates whether trait change is concentrated in specific regions of the phylogeny

-This would happen if changes in one character facilitate changes in another

This allows testing of *directional* hypotheses: "Does aposematism facilitate social (gregarious) behavior?"

-Motivation: if one state (e.g., aposematism) is common, then we expect gregariousness to evolve in its presence more often just by chance. Thus we must account for it.

Discrete Data: Concentrated Changes

 H_0 : Gains/losses in trait 1 are randomly distributed across phylogeny H_1 : Changes in trait 1 depend on changes in trait 2

Procedure:

-Map traits to phylogeny & estimate ancestral states

-Identify locations of changes in each trait

-Calculate:

$$Pr = A/B$$

A: $#(G,L_{obs.trt1}|$ distribution of trait 2, τ) B: $#(G,L_{obs.trt1}| \tau)$

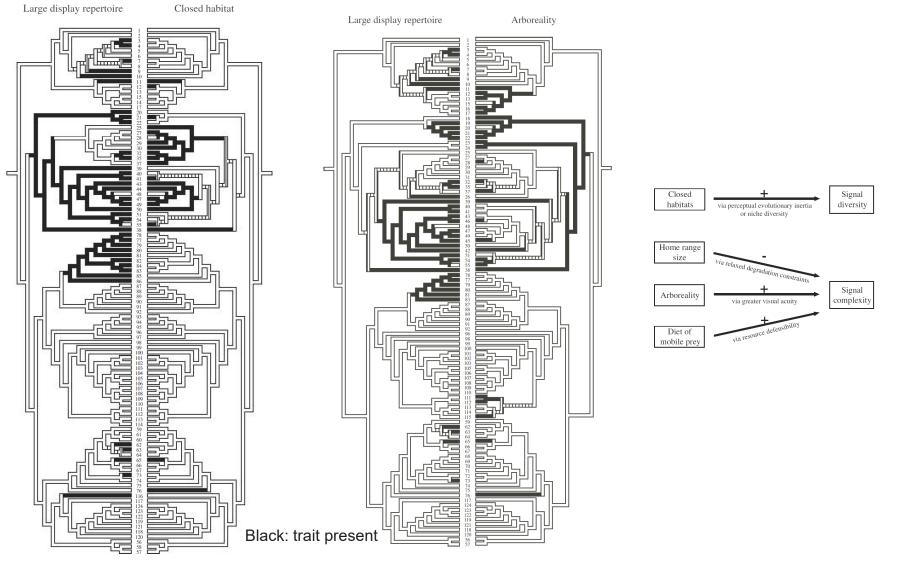
The number of ways gains/losses could occur on phylogeny (B), versus the number of ways G/L occur while accounting for distribution of trait 2 (A)
Calculations are a bit involved, but straightforward
Use probability rules for A & B (the <u>OR</u> rule: for details see Maddison 1990)!

 τ : the phylogeny

Maddison (1990)

Discrete Data: Concentrated Changes

Is signaling behavior associated with ecology in lizards?



Ecology facilitated evolution of increased signal complexity_{Ord et al. (2012)}

Discrete Data: A General Transition Model

The problem can be modeled statistically

-Estimate probability transition matrix between states (P)

$$\mathbf{P}(t) = \begin{bmatrix} p_{00} & p_{01} \\ p_{10} & p_{11} \end{bmatrix}$$

- p_{ij} is probability of transition i \rightarrow j at time t
-Rows sum to 1 (so $p_{00} = (1 - p_{01})$

Looks like P matrices in phylogenetics! (and in fact is derived from that formulation)

More useful to consider transition *RATES*: $P(t) = e^{Q(t)}$

$$\mathbf{Q}(t) = \begin{bmatrix} (1 - q_{01})dt & q_{01}dt \\ q_{10}dt & (1 - q_{10})dt \end{bmatrix}$$

Q is estimated using maximum likelihood (or Bayesian methods) $\mathcal{L}(\mathbf{Q}) = \Pr(\mathbf{X}|\mathbf{Q}, \mathbf{\tau})$

Discrete Data: A General Transition Model

Correlated change via expanded Q matrix

Q describes transitions in both traits across the phylogeny

Hypotheses of trait evolution evaluated by comparing different q-values

Discrete Data: A General Transition Model

Hypothesis testing via model comparison:

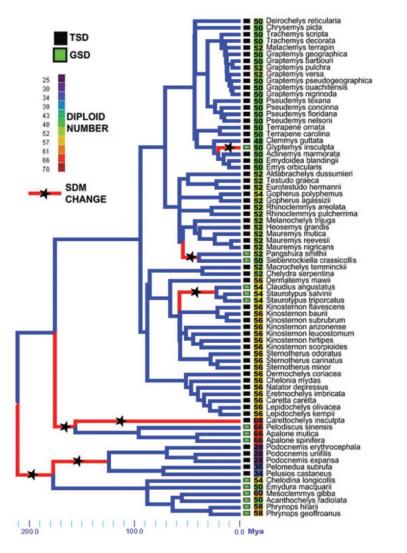
- 1: Jointly estimate $\mathbf{Q} \& \mathcal{L}(\mathbf{Q})$. This is the unconstrained model
- 2: Define 'constrained' model based on biology
 - -These define dependent state changes: $\mathbf{Q}_{\mathbf{d}}$
 - -e.g., G/L rates identical $(q_{13} = q_{31})$;

test	hypothesis	description	statistic
omnibus χ^2 (4 d.f.)	L(I) < L(D)	correlated evolution	L(I) versus $L(D)$
contingent change (1 d.f.)	$q_{12} \neq q_{34}$	change in Y depends upon state of X	$L(D_2)$ versus $L(D_8)$
	$q_{21} \neq q_{43}$	change in Y depends upon state of X	$L(D_7)$ versus $L(D_8)$
	$q_{13} \neq q_{24}$	change in X depends upon state of Y	$L(D_7)$ versus $L(D_8)$
	$q_{31} \neq q_{42}$	change in X depends upon state of Y	$L(D_7)$ versus $L(D_8)$
temporal order or relative rate (1 d.f.)	$q_{12} \neq q_{13}$	order of acquisition of X versus Y	$L(D_7)$ versus $L(D_8)$
	$q_{42} \neq q_{43}$	order of loss of X versus Y	$L(D_7)$ versus $L(D_8)$
alternative models	$q_{ii} = 0$	q_{ii} transitions excluded	$L(D_7)$ versus $L(D_8)$
branch transformation	$\kappa \neq 1$	transformation of branch lengths	$L(D_n)$ versus $L(D_{n+1})$

3: Jointly estimate $\mathbf{Q}_{\mathbf{d}} \& \mathcal{L}(\mathbf{Q}_{\mathbf{d}})$ 4: Model comparison with LRT: $X^2 = -2\log(\mathcal{L}(\mathbf{Q})/\mathcal{L}(\mathbf{Q}_{\mathbf{d}})) *$

Discrete Data: Transition Model

Does sex-determination co-evolve with chromosome changes in turtles?



Did changes in SDM precede Chrom or vice-versa?

> SDM: TSD/GSD Chrom: >52/ <52*

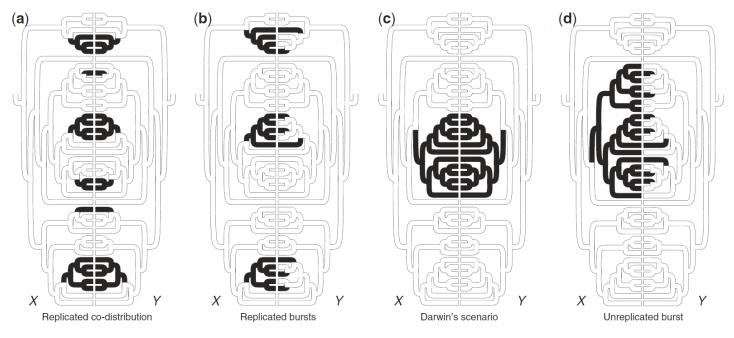
 \mathbf{Q}_{d} : equal transition rates for SDM/Chrom

*q*₁₂=*q*₁₃; *q*₄₂=*q*₄₃

-Changes in traits were associated -No support for one preceding the other

Discrete Data: The Problem of replication

Discrete trait methods sensitive to number of transitions on phylogeny*



C&D lead to inference problems

CAREFUL in interpretation!

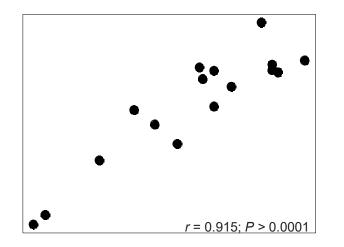
Always plot traits/transitions (don't just examine significance levels)

*We will see same issue with BiSSE models

Maddison & FitzJohn (2015)

Continuous Data: The Problem

Similar issues arise when evaluating association of continuous traits

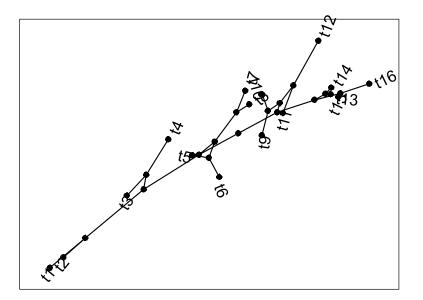


Say we have this correlation,

Here is the same pattern with the phylogeny superimposed

Clearly, closely related taxa are similar.

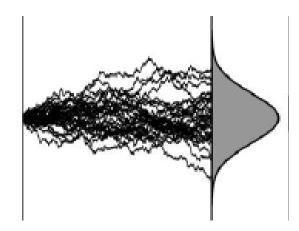
How can we account for this?



Accounting for non-independence due to phylogeny requires a *null model* of evolutionary change

Brownian motion is the null model:

Trait changes are independent from time step to time step Outcome: no change in μ , but $\sigma_y^2 \uparrow \propto$ time



Side-note: this is the continuous-trait model equivalent of the Markov process we discussed earlier

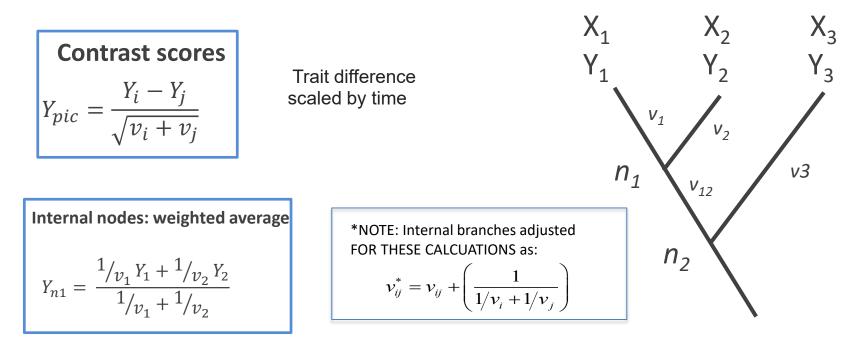
Given this *model*, one can calculate contrast scores between taxa that are independent of one another relative to the phylogeny

Felsenstein (1985)

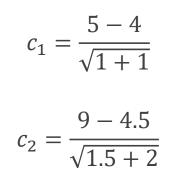
Phylogenetically Independent Contrasts (PIC)

Algorithm:

- -Calculate contrasts for sister taxa
- -Estimate ancestral value
- -Continue recursively down phylogeny

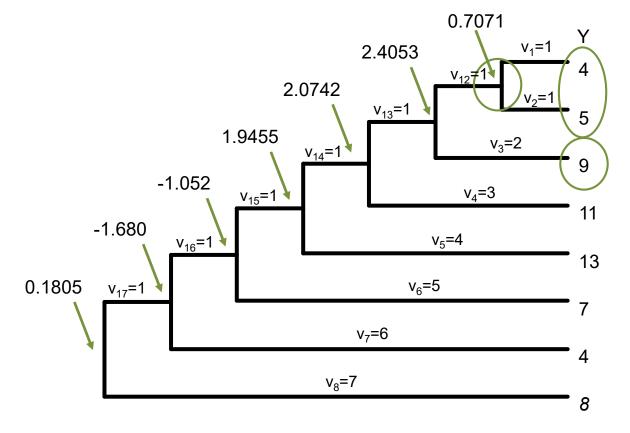


What is this??? The pruning algorithm!!!!



Why? The pruning algorithm!

(need ancestral estimate, which is based on weighted average and rescaled branch lengths)



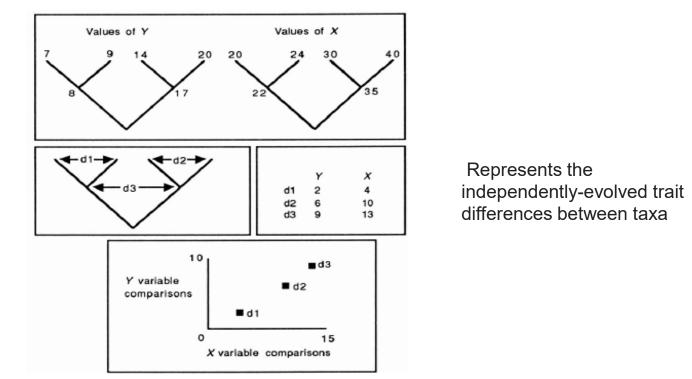
Internal nodes: weighted average

$$Y_{n1} = \frac{1/v_1 Y_1 + 1/v_2 Y_2}{1/v_1 + 1/v_2}$$

NOTE: Internal branches adjusted FOR THESE CALCUATIONS as: $v_{ij}^{} = v_{ij} + \left(\frac{1}{1/v_{i} + 1/v_{j}}\right)$

What are the contrasts?

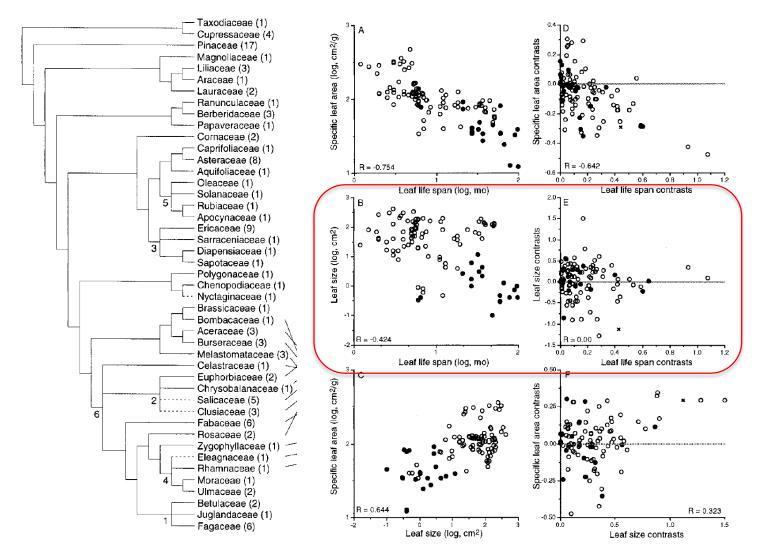
-Amount of evolutionary change since common ancestor



-There are *N*-1 contrasts for *N* taxa

Statistical analysis (regression) performed on contrasts rather than tips data $\hat{\beta} = (X_{pic}^t X_{pic})^{-1} X_{pic}^t Y_{pic}$ (note: through origin because order of tips in contrast irrelevant) Felsenstein (1985)

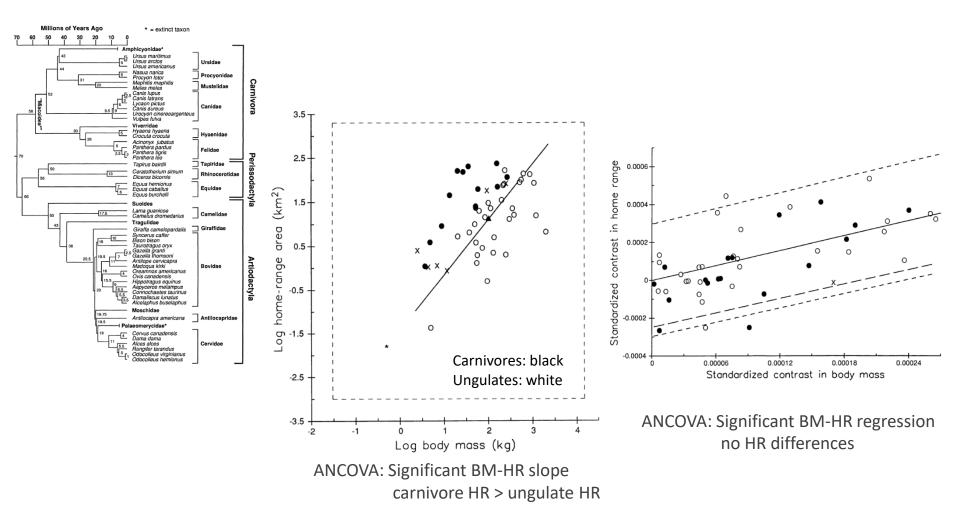
How are leaf traits associated across taxa (prior non-PCM analyses showed correlations)?



Non-phylogenetic association due to angiospermgymnosperm differences

Ackerly and Reich (1999)

Is there an evolutionary relationship between body size and range in mammals?



Conclusions

- -PICs account for non-independence due to phylogeny
- -Clever use of pruning algorithm*
- -Can work with multifurcations (polytomies)

Unanswered questions

- -Is it general?
- -How to examine ANOVA models? Factorial models?
- -What is the relationship between PIC and general statistical theory?

-Stay tuned!

*For a fascinating geometric interpretation and matrix-algebra way of obtaining PICs see Rohlf (2001)